L Number	Hits	Search Text	DB	Time stamp
1	37280	mass adj spectro\$	USPAT; US-PGPUB;	2002/10/15 08:48
2	23	2de same gel	EPO; DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:49
3	14	(mass adj spectro\$) and (2de same gel)	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
4	896	two adj dimensional adj gel	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:58
5	14	(mass adj spectro\$) and (2de same gel)	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:59
6	215	(mass adj spectro\$) and (two adj dimensional adj gel)	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:59
8	9891	hapten	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:59
9	11	((mass adj spectro\$) same (two adj dimensional adj gel)) and hapten	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 08:59
7	90	(mass adj spectro\$) same (two adj dimensional adj gel)	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
10	399	superdex adj "75"	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
11	4	((mass adj spectro\$) and (two adj dimensional adj gel)) and (superdex adj "75")	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 10:43
12	2018976	proteome (w) analysis	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 10:43
13	560	proteomic\$	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 10:43
14	63647	urin\$	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
15	7435	(proteome (w) analysis) same urin\$	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
16	56	proteome adj analysis	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15 10:44
17	0	urin\$ same (proteome adj analysis)	DERWENT USPAT; US-PGPUB; EPO;	2002/10/15
			DERWENT	

18	5	proteomic\$ same urin\$	USPAT;	2002/10/15
10	]	procedure dring	US-PGPUB;	10:45
			EPO;	10.10
			DERWENT	
20	0	(two adj dimensional adj gel) and ((mass	USPAT;	2002/10/15
20		adj spectro\$) and (proteomic\$ same	US-PGPUB;	10:45
i		urin\$))	EPO;	10.45
		urins))	DERWENT	
	o	/day add 117511\ and //magg add	USPAT;	2002/10/15
21	0	, / <del>-</del>		10:45
		spectro\$) and (proteomic\$ same urin\$))	US-PGPUB;	10:45
			EPO;	
	_		DERWENT	0000/10/15
19	3	( (	USPAT;	2002/10/15
	]	urin\$)	US-PGPUB;	10:49
			EPO;	
			DERWENT	
22	119	proteomic\$ and urin\$	USPAT;	2002/10/15
			US-PGPUB;	10:52
			EPO;	
			DERWENT	
23	92	(mass adj spectro\$) and (proteomic\$ and	USPAT;	2002/10/15
		urin\$)	US-PGPUB;	10:52
			EPO;	
			DERWENT	
24	57623	electrophores\$	USPAT;	2002/10/15
			US-PGPUB;	10:52
			EPO;	
			DERWENT	
25	82	((mass adj spectro\$) and (proteomic\$ and	USPAT;	2002/10/15
		urin\$)) and electrophores\$	US-PGPUB;	10:53
			EPO;	
			DERWENT	
26	0	(superdex adj "75") and (((mass adj	USPAT;	2002/10/15
		spectro\$) and (proteomic\$ and urin\$)) and	US-PGPUB;	10:53
		electrophores\$)	EPO;	
			DERWENT	

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FILE 'CAPLUS, MEDLINE, BIOSIS, CA, SCISEARCH, EMBASE' ENTERED AT 10:00:47
· ON 15 OCT 2002
          1806 S PROTEOME (W) ANALYSIS
L2
         10581 S PROTEOMIC?
        518627 S MASS (W) SPECTROMETRY
L3
L4
       1491904 S URIN?
             7 S L1 (S) L4
L5
             3 DUPLICATE REM L5 (4 DUPLICATES REMOVED)
=> s 13 and 16
L7
           2 L3 AND L6
=> d his
     (FILE 'HOME' ENTERED AT 10:00:23 ON 15 OCT 2002)
    FILE 'CAPLUS, MEDLINE, BIOSIS, CA, SCISEARCH, EMBASE' ENTERED AT 10:00:47
    ON 15 OCT 2002
          1806 S PROTEOME (W) ANALYSIS
L1
L2
         10581 S PROTEOMIC?
        518627 S MASS (W) SPECTROMETRY
L3
       1491904 S URIN?
L4
             7 S L1 (S) L4
L5
Lб
             3 DUPLICATE REM L5 (4 DUPLICATES REMOVED)
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1 8- 10

L7

2 S L3 AND L6

=> s 12 (s) 14 L8 95 L2 (S) L4

=> s 13 and 18 L9 38 L3 AND L8 L10 ANSWER 9 OF 14 MEDLINE DUPLICATE 6

TI Peptide mapping of proteins in human body fluids using electrospray ionization Fourier transform ion cyclotron resonance mass spectrometry.

- Human body fluids have been rediscovered in the post-genomic era as great AΒ sources of biological markers and perhaps particularly as sources of potential protein biomarkers of disease. Analytical tools that allow rapid screening, low sample consumption, and accurate protein identification are of great importance in studies of complex biological samples and clinical diagnosis. Mass spectrometry is today one of the most important analytical tools with applications in a wide variety of fields. One of the fastest growing applications is in proteomics, or the study of protein expression in an organism. Mass spectrometry has been used to find post-translational modifications and to identify key functions of proteins in the human body. In this study, we review the use of human body fluids as sources for clinical markers and present new data that show the ability of Fourier transform ion cyclotron resonance (FTICR) mass spectrometry (MS) to identify and characterize proteins in four human body fluids: plasma, cerebrospinal fluid (CSF), saliva, and urine. The body fluids were tryptically digested without any prior separation, purification, or selection, and the digest was introduced into a 9.4 T FTICR mass spectrometer by direct-infusion electrospray ionization (ESI). Even though these samples represent complex biological mixtures, the described method provides information that is comparable with traditional 2D-PAGE data. The sample consumption is extremely low, a few microliters, and the analysis time is only a few minutes. It is, however, evident that the separation of proteins and/or peptides must be included in the methodology, in order to detect low-abundance proteins and other proteins of biological relevance. Copyright 2002 Wiley Periodicals, Inc.
- SO MASS SPECTROMETRY REVIEWS, (2002 Jan-Feb) 21 (1) 2-15. Ref: 54 Journal code: 8219702. ISSN: 0277-7037.
- AU Bergquist Jonas; Palmblad Magnus; Wetterhall Magnus; Hakansson Per;

SWER 11 OF 14 CAPLUS COPYRIGHT 2002 ACS DUPLICATE

- TI Toward **proteomics** in uroscopy: **Urinary** protein profiles after radiocontrast medium administration
- Previous attempts to use urinary protein profiles for diagnostic purposes AΒ have been rather disappointing with respect to their clin. validity, in part because of the insufficient reproducibility, sensitivity, and rapidity of available techniques. Therefore, a newly developed, high-throughput technique, namely surface-enhanced laser desorption/ionization (SELDI) ProteinChip array-time of flight mass spectrometry, was studied, to assess its applicability for protein profiling of urine and to exemplify its use for a group of patients receiving radiocontrast medium. Assessment of the accuracy, sensitivity, and reproducibility of SELDI in test urinary protein profiling was performed. Renal function was studied in 20 male Sprague-Dawley rats before and after i.v. administration of either 1.25 g/kg ioxilan (n = 10) or hypertonic saline soln. (n = 10) as a control. Urine samples from 25 patients undergoing cardiac catheterization were obtained before, immediately after, and 6 to 12 h after the procedure. Administration of ioxilan to rats resulted in changes in the abundance of proteins of 9.9, 18.7, 21.0, and 66.3 kDa. For patients, even in uncomplicated cases of radiocontrast medium infusion during cardiac catheterization, perturbations in the protein compn. occurred but returned to baseline values after 6 to 12 h. Protein with mol. masses of 9.75, 11.75, 23.5, and 66.4 kDa changed in abundance. For patients with impaired renal function, these changes were not reversible within 6 to 12 h. As a proof of principle, one of the peaks, i.e., that at 11.75 kDa, was identified as .beta.2-microglobulin. SELDI is a promising tool for the detection, identification, and characterization of trace amts. of proteins in urine. Even for patients without renal complications, proteins with a broad range of mol. masses either appear in or disappear from the urine. Some of these might represent markers of impending nephropathy.
- SO Journal of the American Society of Nephrology (2001), 12(5), 1026-1035 CODEN: JASNEU; ISSN: 1046-6673
- AU Hampel, Dierk J.; Sansome, Christine; Sha, Ma; Brodsky, Sergey; Lawson, William E.; Goligorsky, Michael S.

L10 ANSWER 2 OF 14 MEDLINE

DUPLICATE 2

TI Proteomic analysis of normal human urinary proteins isolated by acetone precipitation or ultracentrifugation.

- AΒ BACKGROUND: Proteomic techniques have recently become available for large-scale protein analysis. The utility of these techniques in identification of urinary proteins is poorly defined. We constructed a proteome map of normal human urine as a reference protein database by using two differential fractionated techniques to isolate the proteins. METHODS: Proteins were isolated from urine obtained from normal human volunteers by acetone precipitation or ultracentrifugation, separated by two-dimensional polyacrylamide gel electrophoresis (2D-PAGE) and identified by matrix-assisted laser desorption ionization-time-of-flight (MALDI-TOF) mass spectrometry followed by peptide mass fingerprinting. RESULTS: A total of 67 protein forms of 47 unique proteins were identified, including transporters, adhesion molecules, complement, chaperones, receptors, enzymes, serpins, cell signaling proteins and matrix proteins. Acetone precipitated more acidic and hydrophilic proteins, whereas ultracentrifugation fractionated more basic, hydrophobic, and membrane proteins. Bioinformatic analysis predicted glycosylation to be the most common explanation for multiple forms of the same protein. CONCLUSIONS: Combining two differential isolation techniques magnified protein identification from human urine. Proteomic analysis of urinary proteins is a promising tool to study renal physiology and pathophysiology and to determine biomarkers of renal disease.
- SO KIDNEY INTERNATIONAL, (2002 Oct) 62 (4) 1461-9. Journal code: 0323470. ISSN: 0085-2538.
- AU Thongboonkerd Visith; McLeish Kenneth R; Arthur John M; Klein Jon B